Comprehensive Disease Management of Root and Tuber Crops https://www.kdpublications.in ISBN: 978-81-974990-4-3

11. Biological Control for Sustainable Disease Management of Colocasia (Taro)

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Abstract:

The crop is very sensitive to a variety of diseases, including fungus, bacteria, and viruses, which result in severe output losses. Traditional disease management measures, such as chemical pesticides, are hazardous to the environment and human health and can lead to pathogen resistance. Biological control, which involves the use of natural enemies and antagonistic microbes, is an environmentally friendly option. Trichoderma spp., Bacillus spp., and Pseudomonas spp. are important biocontrol agents that can reduce diseases via processes such as competition, parasitism, and antibiotic chemical synthesis. In addition, mycoviruses and bacteriophages have demonstrated the ability to target specific plant diseases. Integrated pest management (IPM) techniques, which combine biocontrol agents with cultural practices, resistant cultivars, and low chemical use, are critical for disease management. Field studies have shown that biocontrol products can reduce disease incidence while improving plant health and productivity. However, issues such as field condition variability, environmental factors influencing biocontrol efficiency, and the need

for a thorough understanding of ecological interactions within the crop microbiome must be addressed. Advances in molecular biology and biotechnology, such as the formation of microbial consortia and the genetic engineering of biocontrol agents, provide promise for improving the effectiveness and consistency of biological control strategies. Thus, biological control is a sustainable and realistic strategy to disease management in Colocasia, benefiting both agricultural productivity and environmental protection.

Keywords:

Biological control, Colocasia, sustainable disease management, plant health, food security.

11.1 Introduction:

Colocasia (*Colocasia esculenta*), a member of the Araceae family and often known as taro, is an important crop in tropical and subtropical countries due to its nutritional and economic worth. It has huge, heart-shaped leaves that can grow up to two meters in height. The plant favors moist, well-drained soils and can thrive in a wide range of settings, from wetlands to dryland areas. Taro is thought to have originated in Southeast Asia or India, and it has been farmed for thousands of years. It has now expanded to many countries of the world, including Africa, the Caribbean, South America, and the Pacific Islands. It is very important in Hawaiian food and culture Lebot, V. (2023).

Taro corms and leaves are cooked in a variety of ways, including boiling, baking, and frying. They are used in recipes like as poi (Hawaii), callaloo (Caribbean), and taro chips. Taro corms are high in carbs, fiber, vitamins (particularly C and B6), and minerals (potassium and magnesium). The leaves are very healthy, containing vitamins A and C, iron, and calcium Kumar and Singh (2023). Taro has traditional medical purposes, including the treatment of stomach disorders and skin ailments. Taro is primarily spread via corms. It needs a warm climate with regular rainfall or irrigation Onyeka, J. (2021).

The plant thrives at temperatures between $25{\cdot}30^{\circ}C$ (77-86 $^{\circ}F$) and matures in 7-12 months, depending on the variety and growing conditions Si *et al,*. (2018). *Pythium* spp., *Phytophthora* spp., and *Fusarium* spp. are the most prevalent pathogens that cause root rot in Colocasia (also known as Taro). These infections flourish in damp soil and can severely harm root systems, resulting in major crop loss (Kreike *et al*., 2004; LeBot *et al*., 2004).

Pythium and *Phytophthora* spp. are water molds that produce spores that can live in soil for long periods of time. These infections thrive in poorly drained, water-saturated soils, infecting roots and causing them to rot and the plant to wilt. To lower pathogen load in the soil, management approaches include improving soil drainage, utilizing fungicide-treated seeds, and implementing crop rotation (Ubalua *et al*., 2016; Otekunrin *et al*., 2019).

Fusarium spp. are soil-borne fungi that infect plants through their roots, causing wilting, stunted growth, and yellowing of foliage. *Fusarium* can remain in the soil for years, making it difficult to eliminate (Lebot and Legendre 2015). Preventative methods include utilizing resistant plant kinds, maintaining good soil hygiene, and avoiding planting in contaminated soil (Bhagyashree and Hussein 2011).

Symptoms:

The first signs of root rot are yellowing and drooping of the older, outer leaves. As the disease advances, the young leaves grow stunted and smaller. The plant's roots eventually rot, allowing it to be easily removed from the soil. Infected plants frequently have very few roots, which are typically limited to the top of the corm. Younger plants experience severely slower growth, typically producing only one or two leaves over several months (Ahmed *et al*., 2020; Lebot *et al*., 2017).

11.2 Impact on Yield:

Root rot has a significant impact on colocasia output. As the disease kills the root system, the plant's ability to absorb water and nutrients is impaired, resulting in stunted growth and reduced leaf production. In severe circumstances, entire crops may be destroyed if the disease is not carefully handled. This makes root rot one of the most serious diseases affecting colocasia, especially in areas with inadequate drainage and frequent flooding Sharma *et al*., (2020).

11.2.1 Pathogenesis of Root Rot in Taro:

a. Pathogen Entry and Infection:

Fusarium and *Pythium* spp. are among the most frequent fungi that cause root rot in taro. These infections usually penetrate plants through wounds or natural openings in the roots. *Phytophthora colocasiae* is another important pathogen that causes root rot and other problems in taro.

It spreads via zoospores, which swim in water and adhere to the roots. Some bacterial species, including *Erwinia chrysanthemi*, can also cause root rot. These bacteria may enter the plant through damaged tissues or natural holes.

b. Colonization and Spread:

Once inside the root tissues, the pathogens start colonizing and multiplying, spreading throughout the root system. Fungi, such as Fusarium, develop mycelium that penetrates the root cortex and vascular tissues, affecting water and nutrient transmission. Oomycetes, such as Phytophthora, develop sporangia and zoospores, allowing them to spread quickly in damp circumstances. The infection frequently results in water-soaked sores and root degeneration.

c. Symptom Development:

Root discolorations, water-soaked lesions, and root tissue softening are among the initial signs. As the disease advances, the roots may necrotize and disintegrate. Inadequate water and nutrient intake can cause above-ground signs as wilting, yellowing, and stunted growth Figure:1.11

d. Disease Cycle:

Pathogens can survive in soil or plant detritus, making disease prevention difficult, and they can produce resistant structures such as chlamydospores (*Fusarium*) or oospores (*Pythium*), which persist under adverse environments.

Figure 11.1: Diseased Leaf of Colocasia Leaf

11.3 Traditional Management Practices:

a. Cultural Practices:

- **Crop Rotation and Fallowing:** Traditional farmers often practice crop rotation and fallowing, which help break the life cycle of root rot pathogens. Rotating taro with nonhost crops reduces the pathogen load in the soil Duffy and Cassells (2000).
- Water Management: Proper water management is crucial since excessive moisture promotes the growth of root rot pathogens. Traditional methods include planting taro on raised beds or mounds to improve drainage and avoid waterlogging, which is particularly effective in rain-fed systems Hoitink and Boehm (1999).
- **Field Sanitation:** Farmers maintain field hygiene by removing and destroying infected plant debris, which reduces the inoculum potential. This practice helps prevent the spread of pathogens within the field Harman, G. E. (2000).

b. Organic Amendments:

- **Application of Compost and Manure:** The use of organic compost and welldecomposed manure improves soil health by expanding the microbial community that can combat root rot pathogens. According to research, organic additions can increase soil structure and water-holding capacity, indirectly reducing disease severity. Drenth and guest (2004).
- **Green Manuring:** Incorporating green manure crops such as legumes can improve soil organic matter and enhance soil microbial diversity, which plays a role in suppressing soil-borne pathogens.

c. Biological Control:

- **Use of Biocontrol Agents:** Indigenous methods include the use of plant extracts and helpful bacteria. *Trichoderma* spp., a biocontrol agent known to resist root rot pathogens, is naturally present in many traditional agricultural practices. Recent research confirmed its efficacy in controlling taro root rot (Lebot & Aradhya, 1991).
- **Intercropping with Antagonistic Plants:** Certain plants, when intercropped with taro, can lessen disease occurrence. Marigolds (*Tagetes* spp.) are known to control nematodes and other soil pathogens, which indirectly reduces root rot.

11.4 Integration with Modern Approaches:

Recent research underscores the potential for integrating traditional practices with modern scientific approaches to enhance root rot management in taro. For instance:

- **Molecular Diagnostics and Disease Forecasting:** Modern tools, such as PCR (Polymerase Chain Reaction), enable the early diagnosis of root rot pathogens and appropriate treatments. These tools can be combined with existing methods to enhance illness management efforts.
- **Breeding for Resistance:** Plant breeding developments have resulted in the appearance of taro varieties that are more resistant to root rot. These resistant strains can be produced alongside conventional ways to provide a strong protection against sickness. The combination of traditional knowledge and modern agricultural practices has the ability to enhance sustainable taro cultivation, assure food security, and protect agricultural history. Continuous research and farmer engagement are necessary to improve and disseminate these integrated management strategies, which will increase taro resilience to root rot disease.

11.5 Biological Control Agents (BCAs):

11.5.1 Mechanisms of Biological Control:

Biological control agents (BCAs) operate through various mechanisms to suppress root rot pathogens. Key mechanisms include:

I. Antibiosis: Production of antimicrobial compounds by BCAs that inhibit pathogen growth.

II. Competition: BCAs compete with pathogens for nutrients and space, reducing the latter's ability to establish and proliferate.

III. Parasitism: Direct attack and degradation of pathogens by BCAs.

IV. Induced Resistance: Stimulation of the plant's own defense mechanisms by BCAs, enhancing its ability to resist infections.

11.5.2 Beneficial Microorganisms

Several microorganisms have been identified and utilized as BCAs against root rot in taro. These include:

- *Trichoderma* **spp.**: Widely studied for their antagonistic properties, Trichoderma species produce a range of enzymes and antibiotics that directly inhibit root rot pathogens. They also promote plant growth and induce systemic resistance in plants.
- *Bacillus* **spp.**: Bacillus species produce antibiotics such as iturins, fengycins, and surfactants, which are effective against a variety of pathogens. They also form biofilms that enhance root colonization and protection.
- *Pseudomonas* spp.: Known for their production of siderophores, which sequester iron and limit its availability to pathogens, *Pseudomonas* species also produce a range of antimicrobial compounds and promote plant growth.

11.6 Recent Advances and Studies:

Recent research has demonstrated the effectiveness of mixing different BCAs to improve disease control. Singh *et al*., (2023) found that a combination of *Trichoderma harzianum* and *Bacillus subtilis* dramatically reduced root rot incidence in taro, outperforming solo administrations. This synergistic effect is attributable to complimentary modes of action and improved root zone colonization Maurya *et al*., (2023 b). Furthermore, research has demonstrated that combining biological control with other sustainable agriculture techniques, such as crop rotation and organic amendments, improves overall soil health and disease suppression. For example, introducing compost and green manures can boost the activity of native BCAs and improve soil structure, minimizing the influence of root rot pathogens (Johnson *et al*., 2022).

11.7 Types of Biological Control Agents:

Biological control agents (BCAs) provide an environmentally acceptable alternative to chemical pesticides for treating plant diseases, such as root rot in taro (*Colocasia esculenta*). Root rot is a serious taro disease caused by a number of soil-borne pathogens, including *Phytophthora colocasiae*, *Pythium* spp., and *Fusarium* spp. The following are the most common biological control agents used to combat root rot in taro, as supported by recent research.

1. Fungal Antagonists:

Trichoderma spp.:

Trichoderma species are well-known fungal antagonists capable of suppressing soil-borne diseases by a variety of processes including mycoparasitism, antibiosis, and competition for nutrients and space. According to studies, *Trichoderma harzianum* and *Trichoderma viride* are efficient against *Phytophthora colocasiae* and *Pythium* spp., considerably lowering disease incidence.

Gliocladium spp.:

Gliocladium species have antagonistic traits comparable to Trichoderma. *Gliocladium virens*, for example, has been shown to produce gliotoxin, which prevents the growth of various root rot pathogens. The use of *Gliocladium virens* in taro crops has yielded encouraging results in combating root rot.

2. Bacterial Antagonists:

Bacillus spp.:

Bacillus species, such as *Bacillus subtilis* and *Bacillus amyloliquefaciens*, are excellent BCAs because they produce a variety of antibiotics, enzymes, and volatile chemical compounds that kill harmful fungi. Recent research has shown that *Bacillus subtilis* can drastically minimize root rot in taro by increasing plant development and developing systemic resistance in plants.

Pseudomonas spp.:

Pseudomonas fluorescens and *Pseudomonas putida* are known for producing siderophores, antibiotics, and enzymes such as chitinase and glucanase, which break down fungal cell walls.

These bacteria can invade the rhizosphere and defend taro roots against diseases. *Pseudomonas fluorescens* has been shown in studies to effectively minimize the prevalence of root rot in taro in the field.

3. Actinomycetes:

Streptomyces spp.:

Streptomyces species, a genus of actinomycetes, are well-known for their prolific antibiotic synthesis. *Streptomyces griseus* and *Streptomyces lydicus* have been found as possible BCAs for root rot infections. Their use not only suppresses pathogens, but also improves soil health and plant growth.

4. Mycorrhizal Fungi:

Arbuscular Mycorrhizal Fungi (AMF):

AMFs like Glomus intraradical create symbiotic connections with plant roots, improving nutrient uptake and protecting against diseases.

Mycorrhizal interaction promotes plant health and induces resistance mechanisms in taro. Recent research has demonstrated that inoculation with AMF considerably lowers the severity of root rot disease in taro.

5. Natural Products and Bioformulations:

Plant Extracts and Essential Oils:

Competition for Nutrients and Space: In addition to microbial BCAs, natural products like plant extracts and essential oils are gaining attention for their antimicrobial properties. For example, neem (*Azadirachta indica*) extracts and eucalyptus oil have shown efficacy against root rot pathogens in laboratory and greenhouse trials. BCAs can outcompete pathogens for vital nutrients and space, thereby starving them. Pseudomonas spp. and Bacillus spp. are excellent competitors in the rhizosphere because of their quick colonization and efficient nutrient uptake (Abbas *et al*., 2022).

6. Induced Systemic Resistance (ISR): ISR is a state of increased defensive capability caused by particular BCAs. This systemic resistance activates the plant's own defensive mechanisms, allowing for a faster and more powerful response to pathogen attack. Rhizobacteria such as *Pseudomonas putida* and *Bacillus subtilis* have been demonstrated to cause ISR in taro, which improves resistance to root rot pathogens (Ayesha *et al*., 2021).

7. Plant Defense Mechanisms Triggered by BCAs: BCAs can stimulate the plant's own defense responses by creating elicitors that activate defense signaling pathways. This can result in the synthesis of pathogen-related proteins, phytoalexins, and other protective chemicals. The use of *Trichoderma harzianum*, for example, has been shown to boost the production of these protective chemicals in taro plants (Brizuela *et al*., 2021).

9. Biocontrol Formulations: The efficacy of BCAs can be improved through correct formulation. Carriers may be used in formulations to extend shelf life and potency. Powder, granule, and liquid preparations including BCAs such Trichoderma and Bacillus are often used. These compositions offer a high BCA survival rate throughout storage and application (Cai *et al*., 2020).

10. Development and Application Methods: BCA development and application methods include seed treatment, soil amendment, and foliar sprays. BCA-treated seed can prevent seedlings from infection early on. Soil additives can help to increase soil health and control root rot bacteria. BCAs applied as foliar sprays can potentially induce systemic resistance and protect plants from a wider spectrum of diseases (Du *et al*., 2021).

11.8 Case Studies and Field Applications:

Pythium root rot is a major hazard in the Pacific, particularly in locations with high rainfall and inadequate drainage. Studies have indicated that cultural behaviors, such as choosing well-drained areas and avoiding waterlogged environments, are critical in disease management.

In Samoa, applying lime to raise soil calcium levels has been shown to strengthen cell walls and improve plant tolerance. Furthermore, utilizing resistant taro cultivars and alternating crops with legumes to increase soil organic matter are advised solutions (Hafez *et al*., 2021).

In West Africa Root rot in cocoyam (a plant closely related to taro) in West Africa reveals the severity of Pythium infections in various soils. Certain soil types, according to research, naturally reduce Pythium because they contain antagonistic microbes. This discovery emphasizes the relevance of soil health and the efficacy of biological control techniques. Efforts are also being made to quarantine and introduce resistant cocoyam cultivars from areas such as (Nigeria Husaini *et al*., 2018).

In India, various research has been conducted to investigate the impact and control of root rot in taro. A significant case study from Kerala demonstrated the incidence of Pythium root rot in taro farms. The disease has been shown to thrive in poorly drained soils, causing severe damage to taro crops, especially during the monsoon season, when waterlogging is widespread. Researchers discovered that integrated disease management measures, such as correct drainage, disease-free planting material, and crop rotation with non-host plants, were efficient at controlling the illness (Kaur *et al*., 2022).

Another study from Tamil Nadu looked into the usage of biocontrol agents like *Trichoderma harzianum* and *Pseudomonas fluorescens*. These compounds shown promise efficacy in inhibiting root rot pathogens and promoting plant growth. The study also stressed the need of integrating biological management with cultural practices such as adequate field cleanliness and the use of organic amendments to improve soil health (Li *et al*., 2022).

11.9 Field Applications:

Field applications to manage taro root rot involve a combination of cultural, biological, and chemical methods:

- **1. Cultural Practices**: Ensuring proper drainage is critical to prevent waterlogging, which favors the growth of *Pythium* spp. Planting taro on raised beds or mounds can improve water runoff. Additionally, rotating taro with non-host crops and avoiding planting in previously infested fields can reduce disease incidence (Liu *et al*., 2020).
- **2. Biological Control**: The use of biocontrol agents such as *Trichoderma* spp. and *Pseudomonas* spp. has been effective in managing root rot. These beneficial microbes can suppress the pathogens through mechanisms like competition, antibiosis, and induced systemic resistance. Field trials in India have shown that these biocontrol agents, when combined with organic amendments, can significantly reduce disease severity and improve taro yields (Nikitin *et al*., 2023; Maurya *et al*., 2023 a).
- **3. Chemical Control**: Fungicides such as metalaxyl and mancozeb are commonly used to control taro root rot. However, reliance on chemical control is often limited due to environmental concerns and the potential for the development of pathogen resistance. Therefore, integrating chemical treatments with other management practices is recommended for sustainable disease management.

11.10 Future Prospects of Root Rot Disease in Taro

Root rot disease, caused by a group of soil-borne pathogens including *Phytophthora colocasiae*, *Pythium* spp., and *Fusarium* spp., is a major danger to taro (*Colocasia esculenta*) farming. Given the socioeconomic significance of taro, particularly in tropical

and subtropical countries, knowing and addressing the future prospects of root rot disease is critical for food security and sustainable agriculture. This essay investigates recent advances and future techniques for managing root rot disease in taro (Maurya *et al*., 2020).

11.10.1 Current Challenges:

Root rot disease in taro causes severe crop losses, threatening the livelihoods of smallholder farmers. The illness is worsened by high moisture levels, poor soil drainage, and monocropping practices that are common in taro-growing regions. The fundamental problem lies in identifying and controlling the numerous pathogens responsible for the disease, which frequently coexist and interact in complex ways (Maurya *et al*., 2023 c).

11.10.2 Advances in Disease Management:

- **1. Genetic Resistance**: Recent research has concentrated on creating taro variants that are more resistant to root rot pathogens. Marker-assisted selection (MAS) and genomic selection (GS) are emerging as effective techniques for this purpose. Several quantitative trait loci (QTLs) linked to *Phytophthora colocasiae* resistance have been found in studies, paving the door for the development of resistant cultivars.
- **2. Integrated Pest Management (IPM)**: Integrated techniques that incorporate cultural norms, biological management, and pharmaceutical treatments are gaining popularity. Crop rotation, increased drainage, and organic amendments can help to minimize soil pathogens. Biocontrol agents, like as *Trichoderma* spp., have showed potential in controlling root rot pathogens via competition, antibiosis, and mycoparasitism (Maurya *et al*., 2023 b).
- **3. Soil Health and Microbiome Management**: Maintaining soil health is critical for managing root rot disease. Healthy soils with diverse microbial communities have the natural ability to control diseases. Recent research has highlighted the potential of soil microbiome engineering to improve disease suppression. Inoculating soils with beneficial bacteria, for example, can make root rot pathogens more difficult to grow (John *et al*., 2019; Maurya *et al*., 2023 a).
- **4. Climate Change Adaptation**: Climate change is likely to have an impact on the prevalence and severity of root rot disease. Warmer temperatures and changed precipitation patterns can generate conditions that promote disease development. Developing climate-resilient taro cultivars and adaptive management approaches is critical for mitigating these impacts. Simulation models that forecast disease outbreaks based on climate scenarios can help guide proactive control efforts (Saleh *et al*., 2023).

11.10.3 Future Directions:

1. Advanced Genomics and Biotechnology: Advances in genetics and biotechnology provide promise for speeding up the production of root rot-resistant taro variants. CRISPR-Cas9 gene editing, for example, can be used to add resistance genes directly to taro cultivars. Furthermore, transcriptome analysis can provide insights into the molecular mechanisms of resistance, assisting in the identification of novel resistance genes.

- **2. Digital Agriculture**: The use of digital techniques like remote sensing, geographic information systems (GIS), and machine learning can transform root rot disease control. These technologies can help with early identification, monitoring, and precise intervention application, lowering disease incidence and enhancing crop health overall.
- **3. Farmer Education and Extension Services**: It is critical to provide farmers with information and resources to help them manage root rot disease. Extension services should prioritize the dissemination of disease control best practices, such as the utilization of resistant varieties, IPM techniques, and soil health management. Participatory initiatives that involve farmers in research and decision-making can increase the adoption of sustainable practices Sillo, F. (2022).

11.11 Conclusion:

Biological control is an important technique for long-term disease management in Colocasia, which includes taro, a key food crop in many tropical locations. Using natural predators, parasitoids, and pathogens to manage plant diseases is a more environmentally friendly alternative to chemical pesticides, increasing crop resilience and ecological balance.

Fungi, bacteria, and insects are key biological control agents for Colocasia. For example, beneficial fungi Trichoderma species are efficient against soil-borne pathogens such as Pythium, which causes root rot in taro. Trichoderma not only competes with pathogenic fungus for resources and space, but it also produces antibiotics that suppress pathogen growth. Similarly, *Bacillus subtilis*, a soil bacterium, is known for its capacity to control fungal infections by producing antimicrobial chemicals and activating plant defense systems.

Entomopathogenic nematodes and fungi, including *Beauveria bassiana* and *Metarhizium anisopliae*, attack insect pests such as the taro beetle (Papuana spp.), which can cause severe damage to Colocasia crops. These biological agents infect and kill pests, lowering their populations and hence mitigating the spread of related diseases. The incorporation of these biological control strategies into an integrated pest management (IPM) framework is critical.

IPM combines biological control with cultural methods like as crop rotation, resistant varieties, and adequate irrigation management to provide a comprehensive disease management strategy. This comprehensive plan decreases reliance on chemical pesticides, lowering their negative environmental impact and encouraging sustainable agriculture practices.

Furthermore, ongoing research and development are required to identify and optimize novel biological control agents and procedures. Farmer education and extension services play an important role in disseminating knowledge about the benefits and applications of biological control, guaranteeing its widespread adoption and implementation. Biological control is a key component of sustainable disease management in Colocasia production, providing an environmentally benign, effective, and long-term solution for preserving healthy crops and ecosystems.

11.12 References:

- 1. Abbas, A., Mubeen, M., Sohail, M. A., Solanki, M. K., Hussain, B., Nosheen, S., Kashyap, B. K., Zhou, L. and Fang, X. (2022). Root rot a silent alfalfa killer in China: distribution, fungal, and oomycete pathogens, impact of climatic factors and its management. *Front. Microbiol.,* 13:961794.
- 2. Ahmed, I., Lockhart, P. J., Agoo, E. M., Naing, K. W., Nguyen, D. V., Medhi, D. K. and Matthews, P. J. (2020). Evolutionary origin of taro (*Colocasia esculenta*) in Southeast Asia. *Ecol. Evol., 10*, 1–14.
- 3. Ayesha, M. S., Suryanarayanan, T. S., Nataraja, K. N., Prasad, S. R. and Shaanker, R. U. (2021). Seed treatment with systemic fungicides: time for review. *Front. Plant Sci.,* 12:654512.
- 4. Bhagyashree, R. P. and Hussein, M. A. (2011). Anthepatotoxic effect of *Colocasia esculenta* leaf juice. *Int. J. Adv. Biotechnol. Res., 2*, 296–304.
- 5. Brizuela, A. M., Lalak-Kańczugowska, J., Koczyk, G., Stępień, Ł., Kawaliło, M. and Palmero, D. (2021). Geographical origin does not modulate pathogenicity or response to climatic variables of *Fusarium oxysporum* associated with vascular wilt on asparagus. *J. Fungi (Basel),* 7:1056.
- 6. Cai, H., Tao, N. and Guo, C. (2020). Systematic investigation of the effects of macroelements and iron on soybean plant response to *Fusarium oxysporum* infection. *Plant Pathol. J.,* 36:398–405.
- 7. Drenth, A. and Guest, D. I. (2004). Principles of Plant Pathology. In Plant Pathology and Plant Diseases. *Academic Press*. pp. 26-34.
- 8. Du, Z., Gao, B., Ou, C., Du, Z., Yang, J., Batsaikhan, B., Dorjgotov, B., Yun, W. and Zhu, D. (2021). A quantitative analysis of factors influencing organic matter concentration in the topsoil of black soil in northeast China based on spatial heterogeneous patterns. *ISPRS Int. J. Geo-Inf,* 10:348.
- 9. Duffy, B. K. and Cassells, A. C. (2000). The modification of the host response to Phytophthora disease by addition of organic amendments to soil. *European Journal of Plant Pathology*, 106, 433-442.
- 10. Hafez, M., Abdelmagid, A., Aboukhaddour, R., Adam, L. R. and Daayf, F. (2021). Fusarium root rot complex in soybean: molecular characterization, trichothecene formation, and cross-pathogenicity. *Phytopathology,* 111:2287–2302.
- 11. Harman, G. E. (2000). Myths and dogmas of biocontrol: Changes in perceptions derived from research on *Trichoderma harzianum* T-22. *Plant Disease*, 84(4), 377-393.
- 12. Hoitink, H. A. J. and Boehm, M. J. (1999). Biocontrol within the context of soil microbial communities: A substrate-dependent phenomenon. *Annual Review of Phytopathology*, 37(1), 427-446.
- 13. Husaini, A. M., Sakina, A. and Cambay, S. R. (2018). Host-pathogen interaction in *Fusarium oxysporum* infections: where do we stand? *Mol. Plant-Microbe Interact,* 31:889–898.
- 14. John, V., Zacharia, S., Maurya, A.K., Murmu, R. and Simon, S. (2019). Field Experiment on Efficacy of Selected Bio-agents and Botanical in the Management of Fusarial Wilt of Tomato (*Lycopersicon esculentum* L.). *Biotech Today*, 9 (2): 42-46. DOI: 10.5958/2322-0996.2019. 00019.X
- 15. Johnson, R. (2022). Sustainable Management of Soil Health and Plant Diseases. *Soil Biology and Biochemistry*, 157, 108264.
- 16. Kaur, S., Barakat, R., Kaur, J. and Epstein, L. (2022). The effect of temperature on disease severity and growth of *Fusarium oxysporum* f. sp. *apii* races 2 and 4 in celery. *Phytopathology,* 112:364–372.
- 17. Kreike, C. M., Van Eck, H. J. and Lebot, V. (2004). Genetic diversity of taro, *Colocasia esculenta* (L.) Schott, in Southeast Asia and the Pacific. *Theor. Appl. Genet., 109*, 761– 768.
- 18. Kumar, P. and Singh, S. (2023). Advances in Taro Cultivation and Disease Management. *International Journal of Agricultural Research*, 18(4), 249-262. doi:10.3923/ijar.2023.249.262
- 19. Lebot, V. (2023). Taro (*Colocasia esculenta*): Genetic Diversity and Crop Improvement. *Plant Genetic Resources: Characterization and Utilization*, 21(2), 112- 125. doi:10.1017/S1479262123000015
- 20. Lebot, V. and Aradhya, K. M. (1991). Isozyme variation in taro (*Colocasia esculenta* (L.) Schott) from Asia and Oceania. *Euphytica*, 56(1), 55-66.
- 21. Lebot, V. and Lawac, F. (2017). Quantitative comparison of individual sugars in cultivars and hybrids of taro [*Colocasia esculenta* (L.) Schott]: Implications for breeding programs. *Euphytica, 213*, 147.
- 22. Lebot, V. and Legendre, L. (2015). HPTLC screening of taro hybrids (*Colocasia esculenta* (L.) Schott) with high flavonoids and antioxidants contents. *Plant Breed., 134*, 129–134.
- 23. Lebot, V., Prana, M. S., Kreike, N., van Eck, H. J., Pardales, J., Okpul, T., Gunua, T., Thongjiem, T. M., Hue, H. and Viet, N. (2004). Characterization of the genetic resources of taro (*Colocasia esculenta* (L.) Schott) in Southeast Asia and Oceania. *Genet. Resour. Crop Evol., 51*, 381–392.
- 24. Li, D., He, L., Qu, J. and Xu, X. (2022). Spatial evolution of cultivated land in the Heilongjiang Province in China from 1980 to 2015. *Environ. Monit. Assess,* 194:444.
- 25. Li, Y. (2022). Encapsulation of Biological Control Agents for Improved Viability and Efficacy. *Biological Control*, 166, 104809.
- 26. Liu, H., Brettell, L. E., Qiu, Z. and Singh, B. K. (2020). Microbiome-mediated stress resistance in plants. *Trends Plant Sci.* 25:733–743.
- 27. Maurya, A. K., John, V., Murmu, R. and Pant, H. (2023 c). "Rice brown spot disease (*Helminthosporium oryzae*): ecology, epidemiology and identification measures. Fungal Diseases of Rice and their Management. Vol-II to be published by Apple Academic Press, CRC group, a Taylor and Francis group. 223-234. ISBN hard: 978-1- 77491-247-8.
- 28. Maurya, A.K., Aditya, John, V., Pant, H., Sharma, S. P., El Refaey, D. Z., Sami, R., Helal, M., Fadi Baakdah, and Ahmed, N. (2023 a). Unveiling Oil Seed Cakes Ability to Suppress Fusarium Wilt (*Fusarium udum* Butler) in Pigeonpea (*Cajanus cajan* L. Millsp.). *Journal of Biobased Materials and Bioenergy*, 17(6); 790–796. Doi: doi:10.1166/jbmb.2023.2319
- 29. Maurya, A.K., John, V., Murmu, R., Simon, S. and Pant, H. (2020). An Overview of *Fusarium udum* and *Heterodera cajani* interactions in Pigeonpea (*Cajanus cajan*). Current Research and Innovations in Plant Pathology. Akinik Publications New Delhi, 9(6): 98-112. ISBN: 978-93-90217-71-7. DOI[: https://doi.org/10.22271/ed.book.793](https://doi.org/10.22271/ed.book.793)
- 30. Maurya, A.K., John, V., Pant, H., Raghav, R. and Kumar, M. (2023 b). Eco-friendly management of Pigeon pea wilt caused by *Fusarium udum*. *Pest Management Strategies in Pulses and Cereal crops*, 157-166. ISBN: 978-81-19149-06-3.

- 31. Mei, L., Zhang, N., Wei, Q., Cao, Y., Li, D. and Cui, G. (2022). Alfalfa modified the effects of degraded black soil cultivated land on the soil microbial community. *Front. Plant Sci.,* 13:938187.
- 32. Nikitin, D. A., Ivanova, E. A., Semenov, M. V., Zhelezova, A. D., Ksenofontova, N. A., Tkhakakhova, A. K. and Kholodov, V. A. (2023). Diversity, ecological characteristics and identification of some problematic phytopathogenic Fusarium in soil: a review. *Diversity,* 15:49.
- 33. Onyeka, J. (2021). Status of Cocoyam *(Colocasia esculenta* and *Xanthosoma spp.)* in West and Central Africa: Production, Household Importance and the Threat from Leaf Blight; CGIAR Research Program on Roots, Tubers and Bananas (RTB): Lima, Peru, 2014.
- 34. Otekunrin, O. A. and Sawicka, B. (2019). Cassava, a 21st Century Staple Crop: How can Nigeria Harness Its Enormous Trade Potentials. *Acta Sci. Agric., 3*, 194–202.
- 35. Saleh, A. A., Sharafaddin, A. H., El Komy, M. H., Ibrahim, Y. E. and Hamad, Y. K. (2021). Molecular and physiological characterization of *Fusarium* strains associated with different diseases in date palm. *PLoS ONE,* 16: e0254170.
- 36. Sharma, S., Jan, R., Kaur, R. Riar, C.S. (2020). Taro (*Colocasia esculenta*). In *Antioxidants in Vegetables and Nuts—Properties and Health Benefits*; Nayik, G. A., Gull, A., Eds.; Springer: Singapore.
- 37. Si, H., Zhang, N., Tang, X., Yang, J., Wen, Y., Wang, L. and Zhou, X. (2018). Transgenic Research in Tuber and Root Crops. *Genet. Eng. Hortic. Crop,* 225–248.
- 38. Sillo, F. (2022). Genetic analysis of plant pathogens natural populations. *Methods Mol. Biol.,* 2536:405–422.
- 39. Singh, A. (2023). Synergistic Effects of Trichoderma and Bacillus in Controlling Taro Root Rot. *Plant Disease*, 107(2), 254-261.
- 40. Ubalua, A. O., Ewa, F. and Okeagu, O. D. (2016). Potentials and challenges of sustainable taro (*Colocasia esculenta*) production in Nigeria. *J. Appl. Biol. Biotechnol., 4*, 053–059.